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### Uptake of dietary RRR- $\alpha$ - and RRR- $\gamma$ -tocopherol by nervous tissues, liver and muscle in vitamin-E-deficient rats

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## Uptake of dietary RRR- $\alpha$ - and RRR- $\gamma$ -tocopherol by nervous tissues, liver and muscle in vitamin-E-deficient rats

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### Abstract

The time course of RRR- $\alpha$ -tocopherol and RRR- $\gamma$ -tocopherol uptake by liver, muscle and selected nervous tissues was studied in vitamin-E-deficient rats fed diets containing either RRR- $\alpha$ -tocopherol or RRR- $\gamma$ -tocopherol over a 60 day period. Feeding rats with a RRR- $\alpha$ -tocopherol-supplemented diet induced in brain, cerebellum, sciatic endoneurium and muscle a marked and regular increase in  $\alpha$ -tocopherol concentration. In addition, the tocopherol concentration in liver reached a plateau very rapidly. In contrast, feeding rats with a diet containing the same level of RRR- $\gamma$ -tocopherol induced a very small increase in  $\gamma$ -tocopherol concentration in brain, cerebellum, sciatic endoneurium and muscle, no change in  $\alpha$ -tocopherol concentration of brain and muscle and a slight but significant decrease in  $\alpha$ -tocopherol concentration in sciatic endoneurium and cerebellum. These results indicate that when  $\gamma$ -tocopherol was supplied continuously in the diet  $\gamma$ -tocopherol accumulated significantly in the tissues but to a much lesser extent than when rats were fed with RRR- $\alpha$ -tocopherol. These results also show that in the tocopherol-deficient rat,  $\gamma$ -tocopherol does not significantly affect the residual  $\alpha$ -tocopherol concentrations in brain or cerebellum, except poorly in sciatic endoneurium.

**Keywords:** Vitamin E; Tocopherol; Uptake; Central nervous system; Peripheral nervous system; Liver

### 1. Introduction

Vitamin E, a major lipid-soluble chain-breaking antioxidant in the membranes, is an important factor in the protection of polyunsaturated fatty acids against peroxidative damage. It is well established that vitamin E deficiency is linked with neurological abnormalities in animals and humans [1–4]. An increasing body of literature indicates that vitamin E plays an important role in membrane function. In addition, specific effects of vitamin E that do not involve its antioxidant function and that act upon the architecture of membranes by controlling their lipid profiles have been reported [5].

Vitamin E is a generic term for all tocopherols and tocotrienols ( $\alpha$ ,  $\beta$ ,  $\gamma$ , and  $\delta$ ) which are found especially in dietary vegetable oils. In France, it has been estimated that vegetable oils supply 60–75% of the dietary requirement for vitamin E [6]. Depending on the vegetable oil used in human diet,  $\gamma$ -tocopherol may represent a significant contribution of the total tocopherols. However, many authors have reported that despite a higher intake of

dietary  $\gamma$ -tocopherol  $\alpha$ -tocopherol is the only antioxidant found in the membranes [7]. Bieri and Poukka-Evarts [8] showed that feeding vitamin-E-depleted rats with a mixture of  $\gamma$ - and  $\alpha$ -tocopherol (ratio 2:1) induces a non-significant reduction of  $\alpha$ -tocopherol and a slight increase in  $\gamma$ -tocopherol concentration of many tissues. In rats fed diets containing low, normal and high levels of tocopherol followed by a single dose of  $\alpha$ -,  $\gamma$ - or both tocopherols, Behrens and Madere [9] showed that plasma  $\gamma$ -tocopherol levels varied inversely with the levels of  $\alpha$ -tocopherol. Recently, Kayden and Traber [10] reported that in humans, the liver is involved in the discrimination between  $\alpha$ - and  $\gamma$ -tocopherol. The  $\alpha$ -tocopherol-binding protein, first described by Catignani and Bieri [15] and responsible for this discrimination was recently purified and characterized by Sato et al. [11,12]. These authors have shown that this protein binds preferentially to the  $\alpha$ -tocopherol. Investigations into discrimination between  $\alpha$ - and  $\gamma$ -tocopherols were carried out either by feeding rats with a mixture of  $\alpha$ - and  $\gamma$ -tocopherol or by dosing laboratory animals or humans with single high dose of  $\alpha$ - or  $\gamma$ -tocopherol [8,13]. However, to our knowledge, until now no data have been reported on the comparative uptake of dietary  $\gamma$ - and  $\alpha$ -tocopherol by liver, muscle and nervous tissues.

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Table 1

Ingredient	g/kg
Casein (delipidated, vitamin-E-free)	220
Sucrose	218.5
Cellulose	20
Corn starch	435
Peanut oil/rapeseed oil 50:50 <sup>a</sup>	50
Mineral mixture <sup>b</sup>	45
Vitamin mixture <sup>c</sup>	10
DL-Methionine	1.5

<sup>a</sup> Tocopherols were eliminated from peanut oil and rapeseed oil by deodorisation (Medeol, Cappelle-la-Grande, France).

<sup>b</sup> Composition of the mineral mixture (per kg of diet): CaHPO<sub>4</sub>, 2H<sub>2</sub>O, 17.1; K<sub>2</sub>HPO<sub>4</sub>, 10.8; CaCO<sub>3</sub>, 8.1; NaCl, 3.2; MgO, 0.9; MgSO<sub>4</sub>, 7H<sub>2</sub>O, 4.0; FeSO<sub>4</sub>, 7H<sub>2</sub>O, 0.39; ZnSO<sub>4</sub>, 7H<sub>2</sub>O, 0.2; MnSO<sub>4</sub>, H<sub>2</sub>O, 0.2; CuSO<sub>4</sub>, 5H<sub>2</sub>O, 0.05; NaF, 0.04; KI, 0.0032; CoCO<sub>3</sub>, 0.001; Na<sub>2</sub>SeO<sub>3</sub>, 5H<sub>2</sub>O, 0.001; (NH<sub>4</sub>)<sub>6</sub>Mo<sub>7</sub>O<sub>24</sub>·4H<sub>2</sub>O, 0.001; CrK(SO<sub>4</sub>)<sub>2</sub>, 12H<sub>2</sub>O, 0.02.

<sup>c</sup> Composition of vitamin supplements (g/kg triturated in dextrose) United States Biochemicals, Cleveland, OH, USA. L-ascorbic acid, 45.0; choline chloride, 75.0; D-calcium pantothenate, 3.0; inositol, 5.0; menadione, 2.25; niacin, 4.5; para-aminobenzoic acid, 5.0; pyridoxine HCl, 1.0; riboflavin, 1.0; thiamine HCl, 1.0; vitamin A acetate, 900 000 IU; calciferol (D2), 100 000 IU; biotine, 20 mg; folic acid, 90 mg; vitamin B<sub>12</sub>, 1.35 mg.

This study was undertaken to examine the kinetics of the dietary uptake of RRR- $\alpha$ - or RRR- $\gamma$ -tocopherols by selected nervous tissues, muscle and liver in tocopherol-deficient rats and thus to compare the respective contribution of the  $\alpha$  and  $\gamma$  forms the diet.

## 2. Materials and methods

Female Sprague-Dawley rats were purchased from Iffa Credo (I'Arbresle, France) and were fed a standard laboratory diet. Animals were maintained under standardized conditions of light (7 am–7 pm), temperature 22°C  $\pm$  1, humidity (70%) and received water ad libitum. From the 14<sup>th</sup> day of gestation, breeding female rats were fed a synthetic vitamin-E-deficient diet (APAE-INRA, Jouy-en-Josas, France) (Table 1). Experimental protocols were approved and meet the guidelines of the governmental agency (Ministry of Agriculture, Authorization n°03007; June 4, 1991).

Rats were maintained on this diet during gestation and suckling and pups were fed the same diet for 8 weeks. At the end of the depletion period, male vitamin-E-deficient rats were divided in two groups and fed the vitamin-E-deficient diet supplemented respectively with RRR- $\alpha$ -tocopherol or RRR- $\gamma$ -tocopherol (0.030 g/kg diet). Six rats of each group were deprived of food for one night and sacrificed under light ether anesthesia at 0, 3, 8, 15, 30, and 60 days. RRR- $\alpha$ -tocopherol and RRR- $\gamma$ -tocopherol were purchased from Eastman (Rochester, NY, USA).

Blood was collected and serum was separated. Muscle, brain and sciatic nerves were rapidly removed. Cerebrum and cerebellum were dissected on a cold plate. The en-

doneurium was isolated by microdissection of the sciatic nerve under a dissecting microscope. All the tissues were lyophilized and stored at –30°C until analysis of tocopherols.

Tocopherols were determined by HPLC in serum and lyophilized tissues according to Ueda and Igarashi [14]. For HPLC determination, a 12.5 cm Lichrosphere RP 18 column containing 4  $\mu$ m particles was used (Merck-Clévenot, Nogent-sur-Marne, France). The eluant, methanol/water 95:5, was pumped at a rate of 1 ml/min with a 2150 LKB pump (Pharmacia LKB, Saint Quentin en Yvelines, France). Tocopherols were detected by their native fluorescence (excitation 295 nm, emission 320 nm) using a fluorometer (Merck-Hitachi, Nogent-sur-Marne, France) to eliminate the UV absorbing compounds that migrate close to  $\alpha$ -tocopherol. The concentrations of  $\alpha$ - and  $\gamma$ -tocopherols were calculated with a 2500 Chromato-Integrator (Merck-Hitachi, Nogent-sur-Marne, France) using  $\delta$ -tocopherol as internal standard. Results are expressed as  $\mu$ g/ml for serum and  $\mu$ g/g fresh weight for tissues except for endoneurium which was expressed as  $\mu$ g/g dry weight (because endoneurium was separated from whole sciatic nerve to avoid contamination by perineurium and fat cells).

Data were expressed as means  $\pm$  S.D. Statistical comparisons of means were made using analysis of variance (ANOVA) with the Bonferroni correction for multiple comparisons. Differences were considered significant at  $P < 0.05$  using the dBSTATS program package (Ashton-Tate, Torrance, USA).

## 3. Results

Table 2 shows the concentrations of  $\alpha$ - and  $\gamma$ -tocopherol in serum, liver, muscle and nervous tissues in rats after 8 weeks of tocopherol deprivation and after 8 weeks of  $\alpha$ - and  $\gamma$ -tocopherol repletion. Feeding rats with a vitamin-E-deficient diet for 8 weeks induced a dramatic reduction in the concentrations of  $\alpha$ -tocopherol in serum, liver, and muscle. In contrast, significant concentrations of  $\alpha$ -tocopherol were observed in all nervous tissue samples from these rats. Curiously, sciatic endoneurium was the only tissue where a significant level of  $\gamma$ -tocopherol was found. Table 2 also shows that the  $\alpha/\gamma$ -tocopherol ratio was higher in forebrain and cerebellum than in the other tissues.

Fig. 1 shows the time course of the concentrations of  $\alpha$ - and  $\gamma$ -tocopherol in the serum of rats fed diets enriched with either RRR- $\alpha$ -tocopherol or RRR- $\gamma$ -tocopherol for 8 weeks. Although the diets supplied the same amounts of tocopherols, the concentrations of  $\alpha$ -tocopherol in serum were higher than those found in serum of rats fed a diet supplemented with RRR- $\gamma$ -tocopherol. Consequently, the time course of the concentrations of  $\alpha$ - and  $\gamma$ -tocopherol in liver displayed a similar pattern (Fig. 2A and 2B).

Table 2

	Time	
	Day-0	Day-60
Serum		
$\alpha$	ND	$4.77 \pm 0.71$
$\gamma$	ND	$1.45 \pm 0.24$
$\alpha/\gamma$		3.30
Liver		
$\alpha$	$0.48 \pm 0.22$	$8.75 \pm 1.35$
$\gamma$	$0.46 \pm 0.06$	$1.48 \pm 0.19$
$\alpha/\gamma$	1.15	5.9
Muscle		
$\alpha$	$0.50 \pm 0.01$	$7.55 \pm 1.13$
$\gamma$	$0.31 \pm 0.07$	$2.00 \pm 0.34$
$\alpha/\gamma$	1.70	3.7
Forebrain		
$\alpha$	$3.75 \pm 0.35$	$14.70 \pm 0.91$
$\gamma$	$0.33 \pm 0.07$	$1.65 \pm 0.20$
$\alpha/\gamma$	11.70	8.9
Cerebellum		
$\alpha$	$2.57 \pm 0.16$	$10.50 \pm 1.31$
$\gamma$	$0.18 \pm 0.01$	$1.30 \pm 0.18$
$\alpha/\gamma$	14.50	8.1
Sciatic endoneurium		
$\alpha$	$9.61 \pm 0.97$	$26.94 \pm 3.41$
$\gamma$	$3.86 \pm 1.02$	$6.42 \pm 0.93$
$\alpha/\gamma$	2.65	4.2

Serum and tissue levels of  $\alpha$ - and  $\gamma$ -tocopherol in rats fed a vitamin-E-deficient diet for 8 weeks (Day-0) and in vitamin-E-deficient rats fed a diet supplemented with either  $\alpha$ - or  $\gamma$ -tocopherol for 8 weeks (Day-60). Values are expressed as  $\mu\text{g}$  of tocopherol/g of fresh tissue except serum ( $\mu\text{g}/\text{ml}$ ) and sciatic endoneurium ( $\mu\text{g}/\text{g}$  dry tissue). Each value represent the mean  $\pm$  S.D. of six animals.

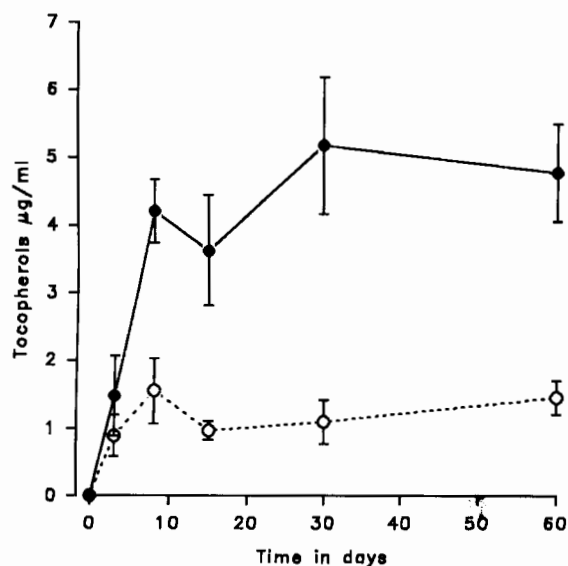


Fig. 1. Kinetics of  $\alpha$ - and  $\gamma$ -tocopherol repletion in serum. Rats previously fed a vitamin-E-deficient diet for 8 weeks were fed for 8 weeks with diets containing either RRR- $\alpha$ -tocopherol (30 mg/kg diet) or RRR- $\gamma$ -tocopherol (30 mg/kg diet). Animals were sacrificed at different times after repletion. Each point represents the mean  $\pm$  S.D. of 6 rats. ●,  $\alpha$ -tocopherol; ○,  $\gamma$ -tocopherol.

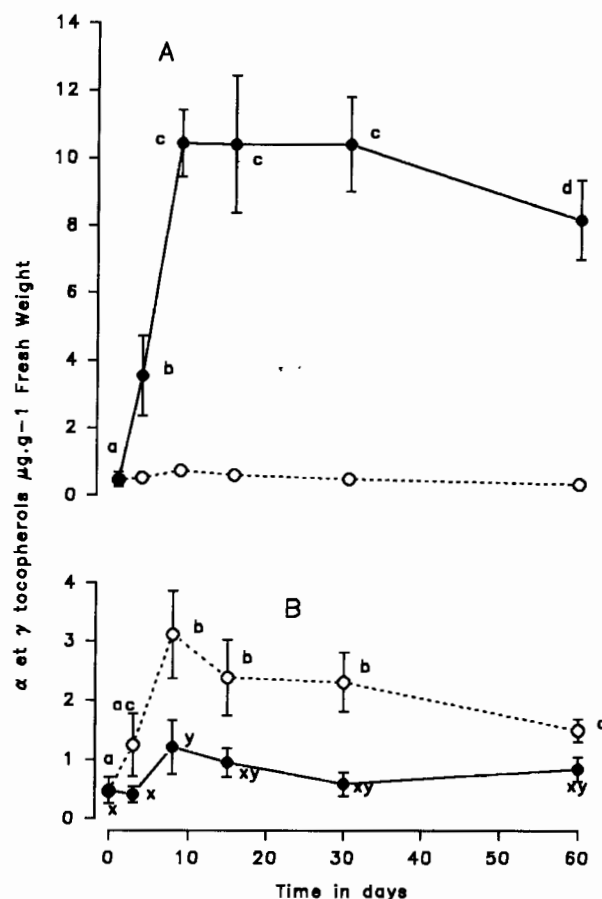


Fig. 2. Kinetics of  $\alpha$ - and  $\gamma$ -tocopherol repletion in liver. Rats previously fed a vitamin-E-deficient diet for 8 weeks were fed for 8 weeks with diets containing either (A) RRR- $\alpha$ -tocopherol (30 mg/kg diet) or (B) RRR- $\gamma$ -tocopherol (30 mg/kg diet). Animals were sacrificed at different times after repletion. Each point represents the mean  $\pm$  S.D. of 6 rats. Values not bearing the same superscript letter are significantly different at  $P < 0.05$ . ●,  $\alpha$ -tocopherol; ○,  $\gamma$ -tocopherol.

In rats fed a diet enriched with RRR- $\alpha$ -tocopherol, the  $\alpha$ -tocopherol concentration in each tissue increased with time and at a different rate. The concentrations of  $\alpha$ -tocopherol in muscle, forebrain and cerebellum increased linearly with time and did not plateau before the end of the experiment (Figs. 3A, 4A and 5A). In contrast, the non-linear increase in the  $\alpha$ -tocopherol concentration in sciatic endoneurium reached a maximum after 2 weeks (Fig. 6A). Liver accumulated  $\alpha$ -tocopherol at a faster rate than any of the other tissues. After only 8 days of repletion with RRR- $\alpha$ -tocopherol,  $\alpha$ -tocopherol concentrations in liver reached a plateau. The concentrations of  $\gamma$ -tocopherol in each tissue were below the limit of detection during the repletion with RRR- $\alpha$ -tocopherol (Figs. 3A, 4A and 5A).

When rats were fed the diet supplemented with RRR- $\gamma$ -tocopherol, significant concentrations of  $\gamma$ -tocopherol were found in all tissues, but these concentrations were much lower than the  $\alpha$ -tocopherol concentrations determined in rats fed the  $\alpha$ -tocopherol supplemented diet (Figs. 2B, 3B, 4B, 5B and 6B). The  $\alpha$ -tocopherol concen-

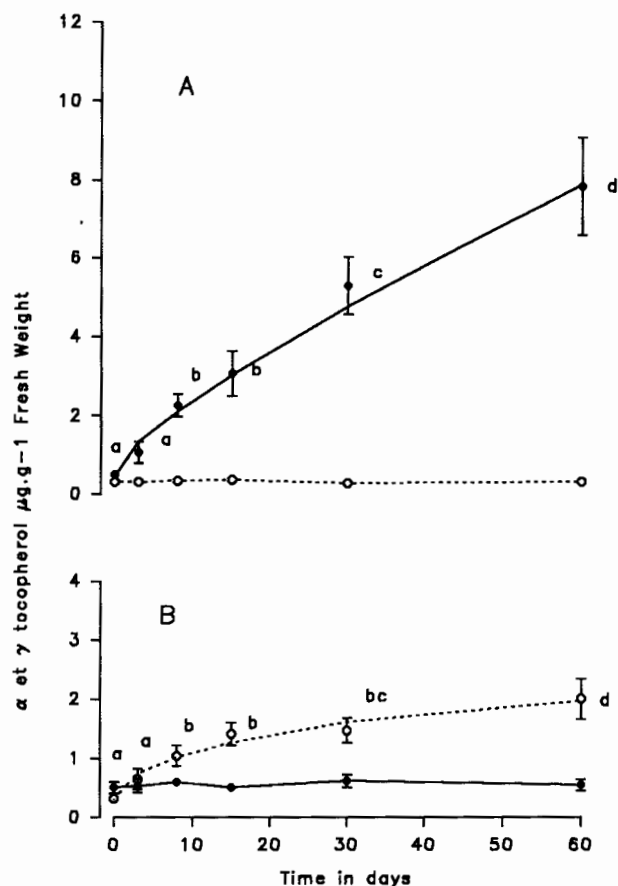


Fig. 3. Kinetics of  $\alpha$ - and  $\gamma$ -tocopherol repletion in muscle. Rats previously fed a vitamin-E-deficient diet for 8 weeks were fed for 8 weeks with diets containing either (A) RRR- $\alpha$ -tocopherol (30 mg/kg diet) or (B) RRR- $\gamma$ -tocopherol (30 mg/kg diet). Animals were sacrificed at different times after repletion. Each point represents the mean  $\pm$  S.D. of 6 rats. Values not bearing the same superscript letter are significantly different at  $P < 0.05$ . ●,  $\alpha$ -tocopherol; ○,  $\gamma$ -tocopherol.

trations in forebrain were maintained for 8 weeks although the diet enriched with  $\gamma$ -tocopherol was devoid of  $\alpha$ -tocopherol (Fig. 4B). Furthermore, the  $\gamma$ -tocopherol concentration in forebrain did not reach the residual  $\alpha$ -tocopherol level even after 8 weeks of repletion with a diet enriched with RRR- $\gamma$ -tocopherol (Fig. 4B). In contrast, the  $\alpha$ -tocopherol concentration decreased significantly in cerebellum and sciatic endoneurium and  $\gamma$ -tocopherol increased very slightly (Figs. 5B and 6B).

#### 4. Discussion

This study demonstrates that when RRR- $\gamma$ -tocopherol was provided continuously in the diet, RRR- $\gamma$ -tocopherol accumulated significantly in the tissues but to a much lesser extent than when rats were fed a RRR- $\alpha$ -tocopherol diet. Furthermore, marked differences in the tissue uptakes were observed.

In 1977, Catigniani and Bieri demonstrated that rat liver cytosol contains a protein that binds  $\alpha$ -tocopherol with high affinity and specificity [15]. In recent years many authors have emphasized the role of this  $\alpha$ -tocopherol binding protein in the transfer of  $\alpha$ -tocopherol between membranes [15–17]. It was shown by Berhens and Madère [9] that the uptake of  $\gamma$ -tocopherol varies inversely with the levels of  $\alpha$ -tocopherol. In addition, Traber et al. [13] have reported that  $\alpha$ -tocopherol as compared with  $\gamma$ -tocopherol is preferentially incorporated and secreted in lipoproteins. In 1987, Ingold et al. [19] reported the synthesis and utilization of deuterium-labeled  $\alpha$ -tocopherol to examine the uptake of natural RRR- $\alpha$ -tocopherol and the biodiscrimination between RRR- $\alpha$ - and SRR- $\alpha$ -tocopherol in various rat tissues under normal dietary conditions. Then, using these deuterium-substituted tocopherols, Traber et al. [20] have shown that the nascent very low density lipoproteins of monkeys fed a mixture of RRR- $\alpha$ - and SRR- $\alpha$ -tocopherol have higher levels of RRR- $\alpha$  than SRR- $\alpha$ -tocopherol. All these results suggest that  $\alpha$ -

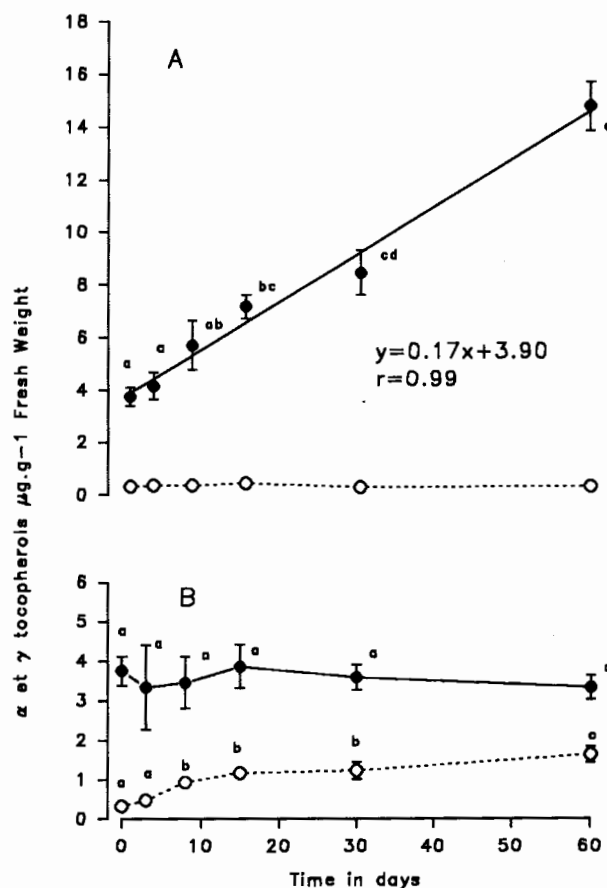


Fig. 4. Kinetics of  $\alpha$ - and  $\gamma$ -tocopherol repletion in forebrain. Rats previously fed a vitamin-E-deficient diet for 8 weeks were fed for 8 weeks with diets containing either (A) RRR- $\alpha$ -tocopherol (30 mg/kg diet) or (B) RRR- $\gamma$ -tocopherol (30 mg/kg diet). Animals were sacrificed at different times after repletion. Each point represents the mean  $\pm$  S.D. of 6 rats. Values not bearing the same superscript letter are significantly different at  $P < 0.05$ . ●,  $\alpha$ -tocopherol; ○,  $\gamma$ -tocopherol.

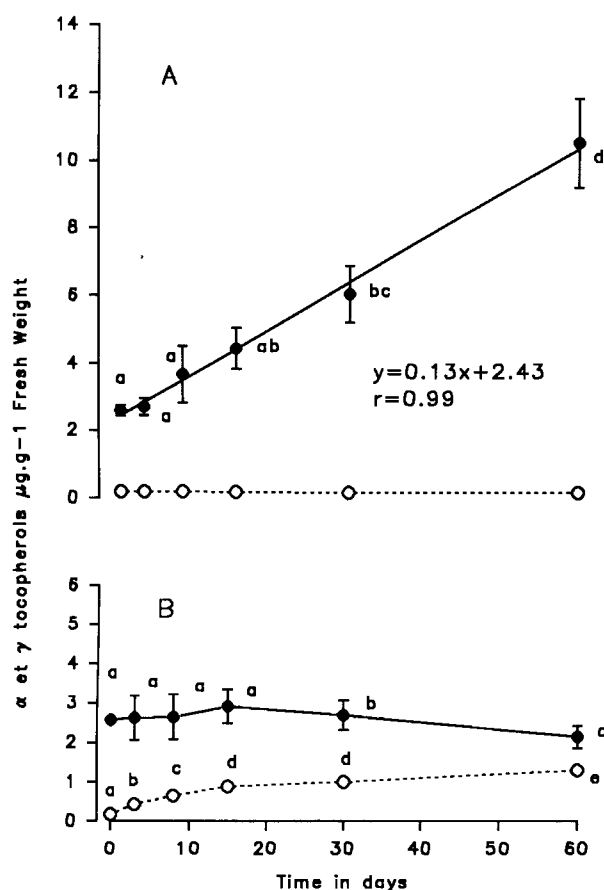


Fig. 5. Kinetics of  $\alpha$ - and  $\gamma$ -tocopherol repletion in cerebellum. Rats previously fed a vitamin-E-deficient diet for 8 weeks, were fed for 8 weeks with diets containing either (A) RRR- $\alpha$ -tocopherol (30 mg/kg diet) or (B) RRR- $\gamma$ -tocopherol (30 mg/kg diet). Animals were sacrificed at different times after repletion. Each point represents the mean  $\pm$  S.D. of 6 rats. Values not bearing the same superscript letter are significantly different at  $P < 0.05$ . ●,  $\alpha$ -tocopherol; ○,  $\gamma$ -tocopherol.

tocopherol binding protein plays an important role in the biodiscrimination between  $\alpha$  and  $\gamma$ -tocopherol and between the C2 epimers of  $\alpha$ -tocopherol. It is well accepted that  $\alpha$ - and  $\gamma$ -tocopherols are equally absorbed by the intestine and secreted in chylomicrons at a similar rate [8,10,21]. Then, following uptake by liver the RRR- $\alpha$  is preferentially incorporated into nascent very low density lipoproteins [20], while the  $\gamma$ -tocopherol is eliminated in the bile [13]. This suggests that the discrimination between  $\alpha$ - and  $\gamma$ -tocopherol takes place in the liver. Vitamin E deficiency has been described in patients consuming normal dietary amounts of vitamin E without abnormalities in lipid absorption and lipoprotein metabolism. In these patients, using deuterated tocopherols, Traber et al. [22] suggested that the low  $\alpha$ -tocopherol levels were linked to an impairment of hepatic  $\alpha$ -tocopherol binding protein activity.

In our experiments the kinetics of the dietary uptake of RRR- $\alpha$ - and RRR- $\gamma$ -tocopherol were investigated in vita-

min-E-deficient rats because, firstly the  $\alpha$ -tocopherol transfer activity is higher in vitamin-E-deficient rats than in control rats [17,18] and secondly in order to determine accurately the increase in RRR- $\alpha$ - or RRR- $\gamma$ -tocopherol. After 8 weeks of vitamin E deprivation the  $\alpha$ -tocopherol concentrations in serum and tissues were similar to those previously reported by our laboratory [23] and by Vatassery et al. in deprived rats [24].

The low plasma  $\gamma$ -tocopherol levels resulting from chylomicron catabolism and exchange between the lipoproteins and the faster rate of  $\gamma$ -tocopherol decrease in cells reported by Tran et al. [25] may contribute to the low  $\gamma$ -tocopherol concentrations in the tissues.

Our study shows, although the two diets contained the same amount of either  $\alpha$ - or  $\gamma$ -tocopherol, that the  $\gamma$ -tocopherol concentrations in serum and liver were always much lower than the concentration of  $\alpha$ -tocopherol determined at the same time in rats fed the diet supplemented with RRR- $\alpha$ -tocopherol. Our results are in agreement with

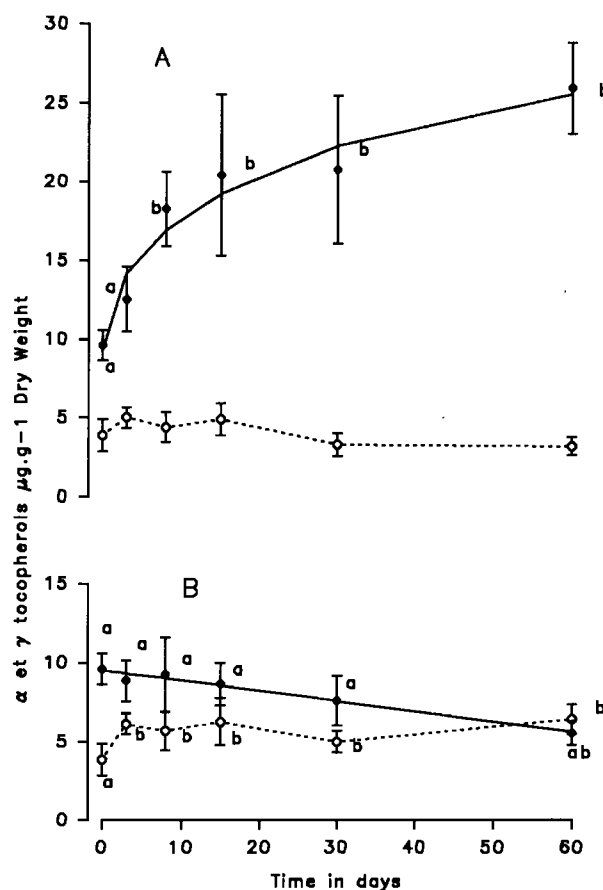


Fig. 6. Kinetics of  $\alpha$ - and  $\gamma$ -tocopherol repletion in sciatic endoneurium. Rats previously fed a vitamin-E-deficient diet for 8 weeks, were fed for 8 weeks with diets containing either (A) RRR- $\alpha$ -tocopherol (30 mg/kg diet) or (B) RRR- $\gamma$ -tocopherol (30 mg/kg diet). Animals were sacrificed at different times after repletion. Each point represents the mean  $\pm$  S.D. of 6 rats. Values not bearing the same superscript letter are significantly different at  $P < 0.05$ . ●,  $\alpha$ -tocopherol; ○,  $\gamma$ -tocopherol.

the data previously reported by Bieri and Poukka Evarts [8] in rats fed a diet containing a mixture of  $\alpha$ - and  $\gamma$ -tocopherol (ratio in the diet 1:2). Our results suggest also that the  $\alpha$ -tocopherol binding protein binds to  $\gamma$ -tocopherol but to a lesser extent than to  $\alpha$ -tocopherol. Determination of the transfer activity of the  $\alpha$ -tocopherol binding protein indicates that  $\gamma$ -tocopherol is a poor substrate [12].

Berhens and Madère [26] reported a linear rate of repletion of  $\alpha$ -tocopherol in liver. On the contrary, our results in liver show a rapid and non-linear kinetic of repletion. The plateau was reached for liver in 8 days. In liver the slope calculated from the linear part of the curve is  $1.25 \mu\text{g}^{-1} \text{day}^{-1}$ . In addition in rats maintained under a normal laboratory diet Ingold et al. [19] reported that the half-life of  $\alpha$ -tocopherol for liver was about 10 days. The last time point of Fig. 2A may indicate that the amount of  $\alpha$ -tocopherol supply by the diet is insufficient to compensate for the requirement of  $\alpha$ -tocopherol needed by growing liver. Of interest was the repletion of the nervous tissues by  $\alpha$ - and  $\gamma$ -tocopherol. The kinetics of repletion of  $\alpha$ -tocopherol in forebrain and cerebellum were linear and no plateau was seen during the experiment. The slopes calculated from the linear regression between RRR- $\alpha$ -tocopherol concentrations over time in forebrain and cerebellum were, respectively,  $0.17 \mu\text{g}^{-1} \text{day}^{-1}$  and  $0.13 \mu\text{g}^{-1} \text{day}^{-1}$ . These results mean that the rate of uptake of  $\alpha$ -tocopherol is similar in forebrain and cerebellum although the  $\alpha$ -tocopherol concentrations in brain and cerebellum are different [7,27]. The capillary endothelial cells of the blood-brain barrier would restrict and regulate the passage of molecules from blood to the brain. The uptake of  $\alpha$ -tocopherol by peripheral tissues may occur via the activity of lipoprotein lipase [28], via the low density lipoprotein receptor or by a non specific uptake [29,30]. The presence of both lipoprotein lipase activity [31] and low density lipoprotein receptors [32] in brain microvessels suggest that the delivery of tocopherol to the brain may involve different mechanisms.

In our experiments,  $\gamma$ -tocopherol was poorly taken up by brain and cerebellum in contrast to  $\alpha$ -tocopherol. Firstly, in rats fed a diet enriched exclusively with  $\gamma$ -tocopherol, the concentration of  $\gamma$ -tocopherol in serum was low, secondly it was reported recently that the uptake in brain consists mainly of RRR- $\alpha$ -tocopherol [19,33], and thirdly that the  $\alpha$ -tocopherol remaining after depletion could inhibit uptake of the  $\gamma$ -tocopherol. Among the nervous tissues, the uptake of  $\alpha$ -tocopherol by the sciatic endoneurium reached a plateau after 2 weeks of repletion and suggests a limiting level of  $\alpha$ -tocopherol repletion.

As in the case of brain and cerebellum, the uptake of  $\alpha$ -tocopherol by muscle was linear. The  $\alpha/\gamma$  ratio in muscle was the highest among the tissues tested.

Finally, these results show that  $\gamma$ -tocopherol was poorly incorporated into tissues but could contribute to antioxidant activity when the diet does not contain  $\alpha$ -tocopherol.

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## References

- [1] Traber, M.G., Sokol, R.J., Ringel, S.P., Neville, H.E., Thellman, C.A. and Kayden H.J. (1987) *N. Engl. J. Med.* 317, 262–265.
- [2] Muller, D.P.R., Llöyd, J.K. and Wolff, O.H. (1983) *Lancet* 29, 225.
- [3] Satya-Murti, S., Howard, L., Krohel, G. and Wolf, B. (1986) *Neurology* 36, 917–921.
- [4] Towfighi, J. (1981) *Acta Neuropathol.* 54, 261–267.
- [5] Buttriss, J.L. and Diplock, A.T. (1988) *Biochim. Biophys. Acta* 962, 81–90.
- [6] Chazan, J.B. (1992) in *Vitamine E, Tocophérols et composés apparentés*. CNERNA-CNRS, pp. 141–157, Polytechnica, Paris.
- [7] Vatassery, G.T., Angerhofer, C.K. and Knox, C.A. (1984) *J. Neurochem.* 43, 409–412.
- [8] Bieri, J.G. and Poukka Evarts, R. (1974) *Am. J. Clin. Nutr.* 27, 980–986.
- [9] Berhens, W.A. and Madère, R. (1987) *J. Nutr.* 117, 1562–1569.
- [10] Kayden, H.J. and Traber, M.G. (1993) *J. Lipid. Res.* 34, 343–358.
- [11] Sato, Y., Hagirawa, K., Arai, H. and Inoue, K. (1991) *FEBS Lett.* 288, 41–45.
- [12] Sato, Y., Arai, H., Miyata, A., Tokita, S., Yamamoto, K., Tanabe, T. and Inoue, K. (1993) *J. Biol. Chem.* 268, 17705–17710.
- [13] Traber, M.G. and Kayden, H.J. (1989) *Am. J. Clin. Nutr.* 49, 517–526.
- [14] Ueda, T. and Igarashi, O. (1990) *J. Micronutr. Anal.* 7, 79–86.
- [15] Catignani, G.L. and Bieri, J.B. (1977) *Biochim. Biophys. Acta* 497, 349–357.
- [16] Murphy, D.J. and Mavis, R.D. (1981) *J. Biol. Chem.* 256, 10464–10468.
- [17] Berhens, W.A. and Madère, R. (1982) *Nutr. Rep. Int.* 25, 1078–1112.
- [18] Mowri, H., Nakagawa, Y., Inoue, K. and Nojima, S. (1981) *Eur. J. Biochemistry* 117, 537–542.
- [19] Ingold, K.U., Burton, G.W., Foster, D.O., Hughes, L., Lindsay, D.A. and Webb, A. (1987) *Lipids* 22, 163–172.
- [20] Traber, M.G., Rudel, L.L., Burton, G.W., Hughes, L., Ingold, K.U. and Kayden, H.J. (1990) *J. Lipid. Res.* 31, 687–694.
- [21] Peake, I.R., Windmuller, H.G. and Bieri, J.G. (1972) *Biochim. Biophys. Acta* 260, 679–688.
- [22] Traber, M.G., Sokol, R.J., Kohlschütter, A., Yokota, T., Muller, D.P.R., Dufour, R. and Kayden, H.J. (1993) *J. Lipid Res.* 34, 201–210.
- [23] Clément, M. and Bourre, J.M. (1993) *Neurosci. Lett.* 164, 163–166.
- [24] Vatassery, G.T., Brin, M.F., Fahn, S., Kayden, H.J. and Traber, M.G. (1988) *J. Neurochem.* 51, 621–623.
- [25] Tran, K. and Chan, A.C. (1992) *Lipids* 27, 38–41.
- [26] Berhens, W.A. and Madère, R. (1991) *J. Nutr.* 121, 454–459.
- [27] Bourre, J.-M. and Clément, M. (1991) *J. Nutr.* 121, 1204–1207.
- [28] Traber, M.G., Olivecrona, T. and Kayden, H.J. (1985) *J. Clin. Invest.* 75, 1729–1734.
- [29] Traber, M.G. and Kayden, H.J. (1984) *Am. J. Clin. Nutr.* 40, 747–751.
- [30] Thellman, C.A. and Shirman, R.B. (1985) *J. Nutr.* 115, 1673–1679.
- [31] Brecher, P. and Kuang, H.T. (1979) *J. Lipid. Res.* 20, 464–471.
- [32] Méresse, S., Delbart, C., Fruchart, J.-C. and Cecchelli, R. (1989) *J. Neurochem.* 53, 340–345.
- [33] Nitta, C., Hayashi, K., Ueda, T. and Igarashi, O. (1993) *Biosci. Biotech. Biochem.* 57, 1406–1407.



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